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l1 and l2 ▲

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DB=USPT; PLUR=YES; OP=OR

<u>L7</u>	l6 and l3	2	<u>L7</u>
<u>L6</u>	encoding DNA and l1	120990	<u>L6</u>
<u>L5</u>	l3 and L4	0	<u>L5</u>
<u>L4</u>	jones.in.	11873	<u>L4</u>
<u>L3</u>	l1 and L2	83	<u>L3</u>
<u>L2</u>	stubbs.in.	252	<u>L2</u>
<u>L1</u>	modified green fluorescent protein	903177	<u>L1</u>

END OF SEARCH HISTORY

WEST**End of Result Set**Generate CollectionPrint

L10: Entry 1 of 1

File: USPT

Jul 7, 1998

DOCUMENT-IDENTIFIER: US 5777079 A

**** See image for Certificate of Correction ****

TITLE: Modified green fluorescent proteins

US Patent No. (1):5777079Detailed Description Text (5):

A molecular interpretation is presented in FIG. 2. If the newly translated apoprotein (top left) evades precipitation into inclusion bodies, the amino group of Gly 67 might cyclize onto the carbonyl group of Ser 65 to form an imidazolidin-5-one, where the process would stop (top center) if O.sub.2 is absent. The new N=C double bond would be expected to promote dehydrogenation to form a conjugated chromophore; imidazolidin-5-ones are indeed known to undergo autoxidative formation of double bonds at the 4-position [Kjaer, A. Acta Chem. Scand. 7, 1030-1035 (1953); Kidwai, A. R. & Devasia, G. M. J. Org. Chem. 27, 4527-4531 (1962)], which is exactly what is necessary to complete the fluorophore (upper right). The protonated and deprotonated species (upper and lower right) may be responsible for the 395 and 470-475 nm excitation peaks, respectively. The excited states of phenols are much more acidic than their ground states, so that emission would come only from a deprotonated species.

Detailed Description Text (7):

According to a first aspect of the present invention, modifications are provided which result in a shift in the ratio of the two excitations peaks of the product after oxidation and cyclization relative to the wild type. Three mutants were found with significant alterations in the ratio of the two main excitation peaks (Table I). The mutations were sequenced and recombined with the wild-type gene in different ways to eliminate neutral mutations and assign the fluorescence effects to single amino acid substitutions, except for H9 where two neighboring mutations have not yet been separated. They all lay in the C terminal part of the protein (Table I), remote in primary sequence from the chromophore formed from residues 65-67.

Detailed Description Text (8):

These and other modifications are defined herein with reference to the amino acid sequence [SEQ ID NO:2] encoded by the reported cDNA [SEQ ID NO:1]; the first amino acid identified is the one found at the indicated location in the reported sequence, while the second indicates the substitution found in the modified form. The fluorescent product derived from a wild-type or modified GFP polypeptide sequence is no longer strictly speaking a simple polypeptide after oxidation and cyclization; however, reference is sometimes made for sake of simplicity herein to the polypeptide (e.g., "wild-type GFP" or "modified GFP") where what is intended would be obvious from the context. Compared with wild-type GFP, H9 (Ser 202.fwdarw.Phe, Thr 203.fwdarw.Ile) had increased fluorescence at 395 nm excitation; P9 (Ile 167.fwdarw.Val) and P11 (Ile 167.fwdarw.Thr) were more fluorescent at 475 nm excitation.

Detailed Description Text (10):

According to another aspect of the invention, a mutant P4 (Tyr 66.fwdarw.His) was identified which was excitable by ultraviolet and fluoresced bright blue in contrast to the green of wild type protein. The excitation and emission maxima were

hypsochromically shifted by 14 and 60 nm respectively from those of wild-type GFP. The mutated DNA was sequenced and found to contain five amino acid substitutions, only one of which proved to be critical: replacement of Tyr 66 in the center of the chromophore by His (corresponding to a change in the GFP cDNA sequence [SEQ ID NO:1] at 196-198 from TAT to CAT).

Detailed Description Text (11):

The surprising tolerance for substitution at this key residue prompted further site-directed mutagenesis to Trp and Phe at this position. Trp gave excitation and emission wavelengths intermediate between Tyr and His (Table I) but was only weakly fluorescent, perhaps due to inefficiency of folding or chromophore formation due to steric considerations. Phe gave weak fluorescence with an excitation maximum at 358 nm and an emission maximum at 442 nm. Accordingly, pursuant to this aspect of the invention modified GFP proteins which fluoresce at different wavelengths (preferably, different by at least 10 nm and more preferably, by at least 50 nm) relative to the native protein are provided, for example, those wherein Tyr 66 is replaced by Phe, His or Trp.

Detailed Description Text (13):

In accordance with further embodiments of this aspect of the invention, a first round of mutagenesis to increase the brightness of Y66W yielded M153T/V163A/N212K as additional substitutions. This mutant was subjected to another round of mutagenesis, resulting in two further sets, N146I and I123V/Y145H/H148R (Table II). The quantum efficiency of these mutants is now comparable to wild-type GFP. The clustering of the substitutions in residues 145 to 163 suggest that those residues lie relatively close to the chromophore and that reductions in the size of their side chains might be compensating for the larger size of tryptophan compared to tyrosine.

Detailed Description Text (15):

Mutagenesis of S65T to shift its wavelengths further to the red yielded M153A/K238E (Table II) as the GFP variant with the longest-wavelength excitation maximum yet described, 504 nm vs. 490 nm for S65T. Surprisingly, the emission peak hardly changed (514 nm vs. 511 nm), so that the separation between the excitation and emission peaks (Stokes' shift) is extremely narrow, only 10 nm. This is one of the smallest values reported for any fluorophore in aqueous solution at room temperature. As in the Y66W series, M153 seems to be influential. It is doubtful that K238E is important, because this substitution has been found to be without effect in other mutants.

Detailed Description Text (16):

As would be readily apparent to those working in the field, to provide the desired fluorescent protein it would not be necessary to include the entire sequence of GFP. In particular, minor deletions at either end of the protein sequence are expected to have little or no impact on the fluorescence spectrum of the protein. Therefore, by a mutant or wild-type GFP sequence for purposes of the present invention are contemplated not only the complete polypeptide and oligonucleotide sequences discussed herein, but also functionally-equivalent portions thereof (i.e., portions of the polypeptide sequences which exhibit the desired fluorescence properties and oligonucleotide sequences encoding these polypeptide sequences). For example, whereas the chromophore itself (position 65-67) is obviously crucial, the locations of known neutral mutations suggest that amino acids 76-115 are less critical to the spectroscopic properties of the product. In addition, as would be immediately apparent to those working in the field, the use of various types of fusion sequences which lengthen the resultant protein and serve some functional purpose in the preparation or purification of the protein would also be routine and are contemplated as within the scope of the present invention. For example, it is common practice to add amino acid sequences including a polyhistidine tag to facilitate purification of the product proteins. As such fusions do not significantly alter the salient properties of the molecules comprising same, modified GFPs as described herein including such fusion sequences at either end thereof are also clearly contemplated as within the scope of the present invention.

Detailed Description Text (17):

Similarly, in addition to the specific mutations disclosed herein, it is well understood by those working in the field that in many instances modifications in

particular locations in the polypeptide sequence may have no effect upon the properties of the resultant polypeptide. Unlike the specific mutations described in detail herein, other mutations provide polypeptides which have properties essentially or substantially indistinguishable from those of the specific polypeptides disclosed herein. For example, the following substitutions have been found to be neutral (i.e., have no significant impact on the properties of the product): Lys 3.fwdarw.Arg; Asp 76.fwdarw.Gly; Phe 99.fwdarw.Ile; Asn 105.fwdarw.Ser; Glu 115.fwdarw.Val; Thr 225.fwdarw.Ser; and Lys 238.fwdarw.Glu. These equivalent polypeptides (and oligonucleotide sequences encoding these polypeptides) are also regarded as within the scope of the present invention. In general, the polypeptides and oligonucleotide sequences of the present invention (in addition to containing at least one of the specific mutations identified herein) will be at least about 85 % homologous, more preferably at least about 90% homologous, and most preferably at least about 95% homologous, to the wild-type GFP described herein. Because of the significant difference in properties observed upon introduction of the specified modifications into a GFP sequence, the presence of the specified modifications relative to the corresponding reported sequence for wild-type GFP [SEQ ID NO:2] are regarded as central to the invention.

Detailed Description Text (22):

The chromophore in GFP is well buried inside the rest of the protein, so much of the dimness of the original point mutants was presumably due to steric mismatch between the substituted amino acid and the cavity optimized for tyrosine. The location of the beneficial mutations implies that residues 145-163 are probably close to the chromophore. The M153A/S65T mutant has the longest wavelengths and smallest Stokes' shift of any known fluorescent protein that does not use a cofactor.

Detailed Description Text (30):

Oligonucleotide-directed mutagenesis at the codon for Ser-65 of GFP cDNA was performed by the literature method [Kunkel, T. A. (1985) Proc. Natl. Acad. Sci. USA 82, 488] using the Muta-Gene Phagemid in Vitro Mutagenesis Kit version 2, commercially available from Bio-Rad, Richmond, Calif. The method employs a bacterial host strain deficient for dUTPase (dut) and uracil-N-glycosylase (ung), which results in an occasional substitution of uracil for thymine in newly-synthesized DNA. When the uracil-containing DNA is used as a wild-type template for oligonucleotide-directed in vitro mutagenesis, the complementary (mutant) strand can be synthesized in the presence of deoxynucleotides, ligase and polymerase using the mutagenic oligonucleotide to prime DNA synthesis; the Version 2 kit utilizes unmodified T7 DNA polymerase to synthesize the complementary strand. When the heteroduplex molecule is transformed into a host with an active uracil-N-glycosylase (which cleaves the bond between the uracil base and the ribose molecule, yielding an apyrimidic site), the uracil-containing wild-type strand is inactivated, resulting in an enrichment of the mutant strand.

Detailed Description Text (42):

Excitation spectra were obtained by collecting emission at the respective peak wavelengths and were corrected by a Rhodamine B quantum counter. Emission spectra were likewise measured at the respective excitation peaks and were corrected using factors from the fluorometer manufacturer (Spex Industries, Edison, N.J.). In cleavage experiments emission spectra were recorded at excitation 368 nm. For measuring molar extinction coefficients, 20 to 30 .mu.g of protein were used in 1 ml of PBS pH 7.4. Quantum yields of wild-type GFP, S65T, and P4-1 mutants were estimated by comparison with fluorescein in 0.1 N NaOH as a standard of quantum yield 0.91 [ed. Miller, J. N., Standards in Fluorescence Spectrometry (Chapman and Hall, New York, 1981)]. Mutants P4 and P4-3 were likewise compared to 9-amino-acridine in water (quantum yield 0.98). W2 and W7 were compared to both standards, which fortunately gave concordant results.

Detailed Description Paragraph Table (3):

	SEQUENCE
LISTING (1) GENERAL INFORMATION: (iii) NUMBER OF SEQUENCES: 2 (2) INFORMATION FOR SEQ ID NO:1: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 716 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1..716 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1: ATGAGTAAAGGAGAAGAACTTTTCACTGGAGTTGTCCCAATTCTTGTT48	

MetSerLysGlyGluGluLeuPheThrGlyValValProIleLeuVal 151015
GAATTAGATGGTGATGTTAATGGGCACAAATTTCTGTCAGTGGAGAG96
GluLeuAspGlyAspValAsnGlyHisLysPheSerValSerGlyGlu 202530
GGTGAAGGTGATGCAACATACGGAAACTTACCCTTAAATTTATTTGC144
GlyGluGlyAspAlaThrTyrGlyLysLeuThrLeuLysPheIleCys 354045
ACTACTGGAAACTACCTGTTCCATGGCCAACACTTGTCACACTTTTC192
ThrThrGlyLysLeuProValProTrpProThrLeuValThrThrPhe 505560
TCTTATGGTGTTCATGCTTTTCAAGATACCCAGATCATATGAAACAG240
SerTyrGlyValGlnCysPheSerArgTyrProAspHisMetLysGln 65707580
CATGACTTTTTCAAGAGTGCCATGCCGAAGGTTATGTACAGGAAAGA288
HisAspPhePheLysSerAlaMetProGluGlyTyrValGlnGluArg 859095
ACTATATTTTTCAAGATGACGGGAACACAAGACACGTGCTGAAGTC336
ThrIlePhePheLysAspAspGlyAsnTyrLysThrArgAlaGluVal 100105110
AAGTTTGAAGGTGATACCCTTGTTAATAGAATCGAGTTAAAAGGTATT384
LysPheGluGlyAspThrLeuValAsnArgIleGluLeuLysGlyIle 115120125
GATTTTAAAGAAGATGGAACATTCTTGGACACAAATTGGAATACAAC432
AspPheLysGluAspGlyAsnIleLeuGlyHisLysLeuGluTyrAsn 130135140
TATAACTCACACAATGTATACATCATGGCAGACAAACAAAAGAATGGA480
TyrAsnSerHisAsnValTyrIleMetAlaAspLysGlnLysAsnGly 145150155160
ATCAAAGTTAACTTCAAAATTAGACACAACATTGAAGATGGAAGCGTT528
IleLysValAsnPheLysIleArgHisAsnIleGluAspGlySerVal 165170175
CAACTAGCAGACCATTATCAACAAATACTCCAATTGGCGATGGCCCT576
GlnLeuAlaAspHisTyrGlnGlnAsnThrProIleGlyAspGlyPro 180185190
GTCCTTTTACCAGACAACCATTACCTGTCCACACAATCTGCCCTTTTCG624
ValLeuLeuProAspAsnHisTyrLeuSerThrGlnSerAlaLeuSer 195200205
AAAGATCCCAACGAAAGAGAGACCACATGGTCCTTCTTGAGTTTGT672
LysAspProAsnGluLysArgAspHisMetValLeuLeuGluPheVal 210215220
ACAGCTGCTGGGATTACACATGGCATGGATGAACTATACAAATA716
ThrAlaAlaGlyIleThrHisGlyMetAspGluLeuTyrLys 225230235 (2) INFORMATION FOR SEQ ID
NO:2: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 238 amino acids (B) TYPE: amino acid
(D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID
NO:2: MetSerLysGlyGluGluLeuPheThrGlyValValProIleLeuVal 151015
GluLeuAspGlyAspValAsnGlyHisLysPheSerValSerGlyGlu 202530
GlyGluGlyAspAlaThrTyrGlyLysLeuThrLeuLysPheIleCys 354045
ThrThrGlyLysLeuProValProTrpProThrLeuValThrThrPhe 505560
SerTyrGlyValGlnCysPheSerArgTyrProAspHisMetLysGln 65707580
HisAspPhePheLysSerAlaMetProGluGlyTyrValGlnGluArg 859095
ThrIlePhePheLysAspAspGlyAsnTyrLysThrArgAlaGluVal 100105110
LysPheGluGlyAspThrLeuValAsnArgIleGluLeuLysGlyIle 115120125
AspPheLysGluAspGlyAsnIleLeuGlyHisLysLeuGluTyrAsn 130135140
TyrAsnSerHisAsnValTyrIleMetAlaAspLysGlnLysAsnGly 145150155160
IleLysValAsnPheLysIleArgHisAsnIleGluAspGlySerVal 165170175
GlnLeuAlaAspHisTyrGlnGlnAsnThrProIleGlyAspGlyPro 180185190
ValLeuLeuProAspAsnHisTyrLeuSerThrGlnSerAlaLeuSer 195200205
LysAspProAsnGluLysArgAspHisMetValLeuLeuGluPheVal 210215220
ThrAlaAlaGlyIleThrHisGlyMetAspGluLeuTyrLys 225230235

CLAIMS:

1. A composition of matters comprising:

a fluorescent modified form of an Aequorea wild-type GFP polypeptide,

characterized in that upon oxidation and cyclization of amino acid residues in said fluorescent modified form corresponding to positions 65 to 67 of wild-type GFP polypeptide sequence (SEQ ID NO:2) said fluorescent modified form exhibits a different excitation and/or emission spectrum from a corresponding product of said wild-type GFP polypeptide sequence,

with the proviso that when said fluorescent modified form comprises a mutation at S65, said mutation at S65 is selected from the group consisting of S65A, S65C, S65T, S65L, S65V, and S65I.

14. The composition according to claim 13,

wherein said fluorescent modified comprises a replacement of Ser at a position corresponding to position 65 of said wild-type GFP polypeptide sequence by an amino acid selected from the group consisting of Ala, Cys, Thr, Leu, Val and Ile.

15. The composition according to claim 14,

wherein said amino acid is Cys or Thr.

16. A functional mutant fluorescent protein, comprising:

a protein with an amino acid sequence that differs from an amino acid sequence of an Aequorea wild type green fluorescent protein (SEQ ID NO:2) by at least one amino acid substitution that is at position 65, wherein said at least one substitution is either S65A, S65C, S65T, S65L, S65V, or S65I,

* wherein said functional mutant fluorescent protein has an excitation or emission different from an excitation spectrum or emission spectrum of said Aequorea wild type green fluorescent protein.

17. The functional mutant fluorescent protein of claim 16,

wherein said at least one amino acid substitution that is at position 65 is S65A.

19. The functional mutant fluorescent protein of claim 16,

wherein said at least one amino acid substitution that is at position 65 is S65C.

21. The functional mutant fluorescent protein of claim 16,

wherein said at least one amino acid substitution that is at position 65 is S65T.

23. The functional mutant fluorescent protein of claim 16,

wherein said at least one amino acid substitution that is at position 65 is S65L.

25. The functional mutant fluorescent protein of claim 16,

wherein said at least one amino acid substitution that is at position 65 is S65V.

27. The functional mutant fluorescent protein of claim 16,

wherein said at least one amino acid substitution that is at position 65 is S65I.

31. The functional mutant fluorescent protein of claim 16,

wherein said functional mutant fluorescent protein further comprises an amino acid sequence which targets said protein to the specific cellular locations.

32. A functional mutant fluorescent protein, comprising:

a protein with an amino acid sequence that differs from an amino acid sequence of an Aequorea wild type green fluorescent protein (SEQ ID NO:2) by at least one amino acid substitution that is at position 66, wherein said at least one substitution is either Y66H or Y66W, further wherein said functional mutant fluorescent protein has an excitation or emission different from an excitation spectrum or emission spectrum of said Aequorea wild type green fluorescent protein.

33. The functional mutant fluorescent protein of claim 32,

wherein said at least one amino acid substitution that is at position 66 is Y66W.

35. The functional mutant fluorescent protein of claim 32,

wherein said at least one amino acid substitution that is at position 66 is Y66H.

43. A functional mutant fluorescent protein, comprising:

a protein with an amino acid sequence that differs from an amino acid sequence of an Aequorea wild type green fluorescent protein (SEQ ID NO:2) by at least one amino acid substitution in the region consisting of positions 65 and 66,

wherein said functional mutant fluorescent protein has an excitation or emission different from an excitation spectrum or emission spectrum of said Aequorea wild type green fluorescent protein.

46. The functional mutant fluorescent protein of claim 43,
further comprising an amino acid substitution that is at position 202.

47. The functional mutant fluorescent protein of claim 46,
wherein said amino acid substitution that is at position 202 is S202F.

48. The functional mutant fluorescent protein of claim 43,
further comprising an amino acid substitution that is at position 203.

49. The functional mutant fluorescent protein of claim 48,
wherein said amino acid substitution that is at position 203 is T203I.

50. The functional mutant fluorescent protein of claim 43,
further comprising an amino acid substitution that is at position 167.

51. The functional mutant fluorescent protein of claim 50,
wherein said amino acid substitution that is at position 167 is I167 V or I167T.

52. The functional mutant fluorescent protein of claim 43,
further comprising an amino acid substitution that is at position 153.

53. The functional mutant fluorescent protein of claim 52,
wherein said amino acid substitution that is at position 153 is M153T or M153A.

54. The functional mutant fluorescent protein of claim 43,
further comprising an amino acid substitution that is at position 238.

55. The functional mutant fluorescent protein of claim 54,
wherein said amino acid substitution that is at position 238 is K238E.

56. The functional mutant fluorescent protein of claim 43,
further comprising an amino acid substitution that is at position 145.

57. The functional mutant fluorescent protein of claim 56,
wherein said amino acid substitution that is at position 145 is Y145H or Y145F.

58. The functional mutant fluorescent protein of claim 43,
further comprising an amino acid substitution that is at position 146.

59. The functional mutant fluorescent protein of claim 58,

wherein said amino acid substitution that is at position 146 is N146I.

60. The functional mutant fluorescent protein of claim 43,
further comprising an amino acid substitution that is at position 163 or 148.

61. The functional mutant fluorescent protein of claim 60,
wherein said amino acid substitution that is at position 163 is V163A and said amino acid substitution at position 148 is H148R.

62. The functional fluorescent protein of claim 43,
further comprising an amino acid substitution that is at position 212 or 123.

63. The functional fluorescent protein of claim 62,
wherein said amino acid substitution that is at position 212 is N212K and said amino acid substitution at position 123 is I123V.

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L11

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DATE: Monday, October 13, 2003

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result set*DB=USPT; PLUR=YES; OP=OR*

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<u>L10</u>	l9 and l8	1	<u>L10</u>
<u>L9</u>	amino acid substitution	720866	<u>L9</u>
<u>L8</u>	5777079.pn.	1	<u>L8</u>
<u>L7</u>	l6 and l3	2	<u>L7</u>
<u>L6</u>	encoding DNA and l1	120990	<u>L6</u>
<u>L5</u>	l3 and L4	0	<u>L5</u>
<u>L4</u>	jones.in.	11873	<u>L4</u>
<u>L3</u>	l1 and L2	83	<u>L3</u>
<u>L2</u>	stubbs.in.	252	<u>L2</u>
<u>L1</u>	modified green fluorescent protein	903177	<u>L1</u>

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WEST[Generate Collection](#)[Print](#)**Search Results - Record(s) 1 through 10 of 83 returned.****1. Document ID: US 6602539 B2**

L3: Entry 1 of 83

File: USPT

Aug 5, 2003

US-PAT-NO: 6602539

DOCUMENT-IDENTIFIER: US 6602539 B2

TITLE: Cooked bean product having reduced solids content and improved viscosity

DATE-ISSUED: August 5, 2003

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
<u>Stubbs</u> ; Timothy A.	Gurnee	IL		
Battle; Betsy O.	Schaumburg	IL		
McPherson; Andrew E.	Mt. Prospect	IL		
Mitchell; Christopher J.	Chicago	IL		
Swenson; Bradley J.	Grayslake	IL		

US-CL-CURRENT: 426/634; 426/629[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachment](#) [Claims](#) [Info](#) [Drawings](#) [Image](#)**2. Document ID: US 6572822 B2**

L3: Entry 2 of 83

File: USPT

Jun 3, 2003

US-PAT-NO: 6572822

DOCUMENT-IDENTIFIER: US 6572822 B2

TITLE: Visual blood glucose test strip

DATE-ISSUED: June 3, 2003

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Jurik; Franklin A.	Pleasanton	CA		
<u>Stubbs</u> ; Andrea	Palo Alto	CA		
Dao; Mimi Diemmy	San Jose	CA		
Chang; Carol	San Francisco	CA		

US-CL-CURRENT: 422/56; 422/58, 422/61, 436/170, 436/95[Full](#) [Title](#) [Citation](#) [Front](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachment](#) [Claims](#) [Info](#) [Drawings](#) [Image](#)**3. Document ID: US 6531322 B1**

L3: Entry 3 of 83

File: USPT

Mar 11, 2003

US-PAT-NO: 6531322

DOCUMENT-IDENTIFIER: US 6531322 B1

TITLE: Visual blood glucose test strip

DATE-ISSUED: March 11, 2003

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Jurik; Franklin A.	Pleasanton	CA		
Stubbs; Andrea	Palo Alto	CA		
Dao; Mimi Diemmy	San Jose	CA		
Chang; Carol	San Francisco	CA		

US-CL-CURRENT: 436/95; 422/56, 422/58, 436/170, 436/85

Full	Title	Citation	Front	Referred	Classification	Date	Reference	Sequence	Attachment	Claims	Page	Image	Image
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└ 4. Document ID: US 6522925 B1

L3: Entry 4 of 83

File: USPT

Feb 18, 2003

US-PAT-NO: 6522925

DOCUMENT-IDENTIFIER: US 6522925 B1

TITLE: System and method for detection enhancement programming

DATE-ISSUED: February 18, 2003

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Gilkerson; James O.	Stillwater	MN		
Conley; Vickie L.	Woodbury	MN		
Stubbs; Scott	Maple Grove	MN		
Lang; Douglas J.	Arden Hills	MN		

US-CL-CURRENT: 607/30

Full	Title	Citation	Front	Referred	Classification	Date	Reference	Sequence	Attachment	Claims	Page	Image	Image
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└ 5. Document ID: US 6493579 B1

L3: Entry 5 of 83

File: USPT

Dec 10, 2002

US-PAT-NO: 6493579

DOCUMENT-IDENTIFIER: US 6493579 B1

TITLE: System and method for detection enhancement programming

DATE-ISSUED: December 10, 2002

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Gilkerson; James O.	Stillwater	MN		
Conley; Vickie L.	Woodbury	MN		
Stubbs; Scott	Maple Grove	MN		
Lang; Douglas J.	Arden Hills	MN		

US-CL-CURRENT: 607/5; 607/31

Full	Title	Citation	Print	Review	Classification	Date	Reference	Sequence	Attachment
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Full	Drawings	Image
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└ 6. Document ID: US 6455614 B1

L3: Entry 6 of 83

File: USPT

Sep 24, 2002

US-PAT-NO: 6455614

DOCUMENT-IDENTIFIER: US 6455614 B1

TITLE: Chlorine-free, zero voc, waterborne adhesion promoter for polyolefinic substrates

DATE-ISSUED: September 24, 2002

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Jackson; Michael L.	LaGrange	IL		
Stubbs; Frank A.	Schererville	IN		
Mecozzi; Joseph M.	Hammond	IN		
Miklos; David J.	Berwyn	IL		
Neymark; Alexander L.	Chicago	IL		

US-CL-CURRENT: 523/401; 523/406, 523/407, 523/439, 525/117

Full	Title	Citation	Print	Review	Classification	Date	Reference	Sequence	Attachment
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└ 7. Document ID: US 6447527 B1

L3: Entry 7 of 83

File: USPT

Sep 10, 2002

US-PAT-NO: 6447527

DOCUMENT-IDENTIFIER: US 6447527 B1

**** See image for Certificate of Correction ****

TITLE: Apparatus and methods for the penetration of tissue

DATE-ISSUED: September 10, 2002

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Thompson; Ronald J.	Ft. Thomas	KY	41075	
Stubbs; Jack B.	Waynesville	OH	45068	

US-CL-CURRENT: 606/174

Full	Title	Citation	Print	Review	Classification	Date	Reference	Sequence	Attachment
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8. Document ID: US 6441058 B2

L3: Entry 8 of 83

File: USPT

Aug 27, 2002

US-PAT-NO: 6441058

DOCUMENT-IDENTIFIER: US 6441058 B2

TITLE: Abrasive articles having abrasive layer bond system derived from solid, dry-coated binder precursor particles having a fusible, radiation curable component

DATE-ISSUED: August 27, 2002

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Thurber; Ernest L.	Woodbury	MN		
Larson; Eric G.	Lake Elmo	MN		
Dahlke; Gregg D.	St. Paul	MN		
DeVoe; Robert J.	Oakdale	MN		
Kirk; Alan R.	Cottage Grove	MN		
Meierotto; Mark R.	Hudson	WI		
Stubbs; Roy	Nuneaton			GB

US-CL-CURRENT: 522/96; 522/103, 522/107, 522/170, 522/173, 522/174, 522/175, 522/179, 522/181, 522/182[Full](#) [Title](#) [Citation](#) [Print](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#)[Full](#) [Draw Data](#) [Image](#)

9. Document ID: US 6277160 B1

L3: Entry 9 of 83

File: USPT

Aug 21, 2001

US-PAT-NO: 6277160

DOCUMENT-IDENTIFIER: US 6277160 B1

**** See image for Certificate of Correction ****

TITLE: Abrasive article and method of making such article

DATE-ISSUED: August 21, 2001

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Stubbs; Roy	Nuneaton			GB
Culler; Scott R.	Burnsville	MN		
Liepa; Mara E.	St. Paul	MN		
Bange; Donna W.	Eagan	MN		
Haas; John D.	Roseville	MN		

US-CL-CURRENT: 51/295; 51/293, 51/298, 51/309[Full](#) [Title](#) [Citation](#) [Print](#) [Review](#) [Classification](#) [Date](#) [Reference](#) [Sequences](#) [Attachments](#)[Full](#) [Draw Data](#) [Image](#)

10. Document ID: US 6228133 B1

L3: Entry 10 of 83

File: USPT

May 8, 2001

US-PAT-NO: 6228133

DOCUMENT-IDENTIFIER: US 6228133 B1

**** See image for Certificate of Correction ****

TITLE: Abrasive articles having abrasive layer bond system derived from solid,
dry-coated binder precursor particles having a fusible, radiation curable component

DATE-ISSUED: May 8, 2001

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Thurber; Ernest L.	Woodbury	MN		
Larson; Eric G.	Lake Elmo	MN		
Dahlke; Gregg D.	St. Paul	MN		
DeVoe; Robert J.	Oakdale	MN		
Kirk; Alan R.	Cottage Grove	MN		
Meierotto; Mark R.	Hudson	WI		
Stubbs; Roy	Nuneaton			GB

US-CL-CURRENT: 51/295; 51/297, 51/298[Full](#) | [Title](#) | [Citation](#) | [Front](#) | [Review](#) | [Classification](#) | [Date](#) | [Reference](#) | [Sequence](#) | [Attachments](#)[Find](#) | [Original](#) | [Image](#)[Generate Collection](#)[Print](#)**Terms****Documents**

11 and L2

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WEST[Generate Collection](#)[Print](#)**Search Results - Record(s) 61 through 70 of 83 returned.****61. Document ID: US 5071661 A**

L3: Entry 61 of 83

File: USPT

Dec 10, 1991

US-PAT-NO: 5071661

DOCUMENT-IDENTIFIER: US 5071661 A

TITLE: Process for dehydrating potato products

DATE-ISSUED: December 10, 1991

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
<u>Stubbs</u> ; Clifford A.	Iona	ID		
Willard; Miles J.	Idaho Falls	ID		

US-CL-CURRENT: 426/96; 426/102, 426/272, 426/273, 426/292, 426/293, 426/464,
426/473, 426/637[Full](#) | [Title](#) | [Citation](#) | [Front](#) | [Reverse](#) | [Classification](#) | [Date](#) | [Reference](#) | [Dependent](#) | [Attachment](#)[Full](#) | [Draw Data](#) | [Image](#)**62. Document ID: US 5052410 A**

L3: Entry 62 of 83

File: USPT

Oct 1, 1991

US-PAT-NO: 5052410

DOCUMENT-IDENTIFIER: US 5052410 A

TITLE: Device for controlling eating and smoking habits

DATE-ISSUED: October 1, 1991

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
<u>Stubbs</u> ; James M.	Rockingham	NC	28379	

US-CL-CURRENT: 128/859; 128/860, 604/909[Full](#) | [Title](#) | [Citation](#) | [Front](#) | [Reverse](#) | [Classification](#) | [Date](#) | [Reference](#) | [Sequence](#) | [Attachment](#)[Full](#) | [Draw Data](#) | [Image](#)**63. Document ID: US 5029959 A**

L3: Entry 63 of 83

File: USPT

Jul 9, 1991

US-PAT-NO: 5029959

DOCUMENT-IDENTIFIER: US 5029959 A

**** See image for Certificate of Correction ****

TITLE: Multiple ring guide for payout testing of optical fibers

DATE-ISSUED: July 9, 1991

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Stubbs; Scott F.	Tucson	AZ		

US-CL-CURRENT: 385/134; 242/125, 242/157R, 242/566, 385/123, 57/352, 57/71

Full Title Citation Front Review Classification Date Reference Sequence Attachment

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└ 64. Document ID: US 5003590 A

L3: Entry 64 of 83

File: USPT

Mar 26, 1991

US-PAT-NO: 5003590

DOCUMENT-IDENTIFIER: US 5003590 A

TITLE: Encoding an optical video disc to inhibit video tape recording

DATE-ISSUED: March 26, 1991

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Lechner; Bernard J.	Princeton	NJ		
Stubbs; Graham S.	Poway	CA		
Leonard; Eugene	Sands Point	NY		

US-CL-CURRENT: 380/204; 360/60, 380/214

Full Title Citation Front Review Classification Date Reference Sequence Attachment

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└ 65. Document ID: US 4868785 A

L3: Entry 65 of 83

File: USPT

Sep 19, 1989

US-PAT-NO: 4868785

DOCUMENT-IDENTIFIER: US 4868785 A

**** See image for Certificate of Correction ****

TITLE: Block diagram editor system and method for controlling electronic instruments

DATE-ISSUED: September 19, 1989

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Jordan; Dale A.	Aloha	OR		
Fitzsimmons; Lynne A.	Portland	OR		
Greenseth; William A.	Portland	OR		
Hoffman; Gregory L.	Beaverton	OR		
Stubbs; David D.	Portland	OR		

US-CL-CURRENT: 345/440

Full	Title	Citation	Front	Revised	Classification	Date	Reference	Conferences	Attachment
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└ 66. Document ID: US 4812996 A

L3: Entry 66 of 83

File: USPT

Mar 14, 1989

US-PAT-NO: 4812996

DOCUMENT-IDENTIFIER: US 4812996 A

TITLE: Signal viewing instrumentation control system

DATE-ISSUED: March 14, 1989

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Stubbs; David D.	Portland	OR		

US-CL-CURRENT: 702/123; 324/121R, 324/76.19, 324/76.24, 345/440, 345/440.1, 345/661, 345/808, 345/970

Full	Title	Citation	Front	Revised	Classification	Date	Reference	Conferences	Attachment
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└ 67. Document ID: US 4760797 A

L3: Entry 67 of 83

File: USPT

Aug 2, 1988

US-PAT-NO: 4760797

DOCUMENT-IDENTIFIER: US 4760797 A

TITLE: Method and apparatus for automated tie detection and tamping

DATE-ISSUED: August 2, 1988

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Stubbs; John L.	Charlotte	NC		
Patton; Wesley H.	Atlanta	GA		
Thompson; Jeffrey L.	Lawrenceville	GA		

US-CL-CURRENT: 104/12; 180/168, 246/187B, 701/19, 701/50

Full	Title	Citation	Front	Revised	Classification	Date	Reference	Conferences	Attachment
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└ 68. Document ID: US 4597500 A

L3: Entry 68 of 83

File: USPT

Jul 1, 1986

US-PAT-NO: 4597500

DOCUMENT-IDENTIFIER: US 4597500 A

TITLE: Tamper-resistant closures for containers

DATE-ISSUED: July 1, 1986

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
<u>Stubbs</u> ; Peter	Dartford			GB

US-CL-CURRENT: 215/256; 215/258

Full	Title	Citation	Front	Review	Classification	Date	Reference	Sequence	Attachment
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└ 69. Document ID: US 4582680 A

L3: Entry 69 of 83

File: USPT

Apr 15, 1986

US-PAT-NO: 4582680

DOCUMENT-IDENTIFIER: US 4582680 A

TITLE: Multiphase backing materials for piezoelectric broadband transducers

DATE-ISSUED: April 15, 1986

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Bar-Cohen; Yoseph	Dayton	OH		
<u>Stubbs</u> ; David A.	Waynesville	OH		
Hoppe; Wally C.	Huber Heights	OH		

US-CL-CURRENT: 419/23; 29/25.35, 419/31, 419/32, 419/48, 419/5, 419/60, 419/8

Full	Title	Citation	Front	Review	Classification	Date	Reference	Sequence	Attachment
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└ 70. Document ID: US 4581070 A

L3: Entry 70 of 83

File: USPT

Apr 8, 1986

US-PAT-NO: 4581070

DOCUMENT-IDENTIFIER: US 4581070 A

TITLE: Multiphase backing materials for piezoelectric broadband transducers

DATE-ISSUED: April 8, 1986

INVENTOR-INFORMATION:

NAME	CITY	STATE	ZIP CODE	COUNTRY
Bar-Cohen; Yoseph	Dayton	OH		
<u>Stubbs</u> ; David A.	Waynesville	OH		
Hoppe; Wally C.	Huber Heights	OH		

US-CL-CURRENT: 75/247; 419/23, 419/5, 419/8, 420/555, 420/563, 420/589, 428/546, 428/570, 428/929, 75/228, 75/245

Full	Title	Citation	Front	Review	Classification	Date	Reference	Sequence	Attachment
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